

3,5-Dibenzoyl-4-(3-phenoxyphenyl)-1,4-dihydro-2,6-dimethylpyridine (DP7) as a new multidrug resistance reverting agent devoid of effects on vascular smooth muscle contractility

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1 The aim of this study was to investigate the effects of 3,5-diacetyl- (DP1–DP5) and 3,5-dibenzoyl-1,4-dihydropyridines (DP6–DP11) on vascular functions *in vitro*, by comparing their mechanical and electrophysiological actions in rat aorta rings and single rat tail artery myocytes, respectively, and to quantify their multidrug resistance (MDR)-reversing activity in L5178 Y mouse T-lymphoma cells transfected with MDR1 gene.

2 In rat aorta, the 11 compounds tested, but 3,5-dibenzoyl-4-(3-phenoxyphenyl)-1,4-dihydro-2,6-dimethylpyridine (DP7), 3,5-dibenzoyl-4-(3-chlorophenyl)-1,4-dihydro-2,6-dimethylpyridine (DP9), 3,5-dibenzoyl-4-(4-chlorophenyl)-1,4-dihydro-2,6-dimethylpyridine (DP10) and 3,5-dibenzoyl-4-phenyl-1,4-dihydro-2,6-dimethylpyridine (DP11), antagonized 60 mM K⁺ (K60)-induced contraction in a concentration-dependent manner, with IC₅₀ (M) values ranging between 5.65 × 10^{−7} and 2.23 × 10^{−5}.

3 The 11 dihydropyridines tested, but DP7, inhibited L-type Ca²⁺ current recorded in artery myocytes in a concentration-dependent manner, with IC₅₀ (M) values ranging between 1.12 × 10^{−6} and 6.90 × 10^{−5}.

4 The K⁺-channel opener cromakalim inhibited the Ca²⁺-induced contraction in K30 but not that evoked in K60. On the contrary, DP7 was ineffective in both experimental conditions.

5 When the rings were preincubated with 1 mM Ni²⁺ plus 1 μM nifedipine, the response to phenylephrine was significantly reduced by 2,5-di-*t*-butyl-1,4-benzohydroquinone (BHQ), a well-known endoplasmic reticulum Ca²⁺-ATPase inhibitor. DP7 had no effects on this model system.

6 In L5178 MDR cell line, the 11 dihydropyridines tested, but 3,5-diacetyl-4-(2-nitrophenyl)-1,4-dihydro-2,6-dimethylpyridine (DP1), 3,5-diacetyl-4-(3-phenoxyphenyl)-1,4-dihydro-2,6-dimethylpyridine (DP2) and 3,5-diacetyl-4-(3-chlorophenyl)-1,4-dihydro-2,6-dimethylpyridine (DP4), exhibited an MDR-reversing activity, with IC₅₀ values ranging between 3.02 × 10^{−7} and 4.27 × 10^{−5}, DP7 being the most potent.

7 In conclusion, DP7 may represent a lead compound for the development of potent dihydropyridine MDR chemosensitizers devoid of vascular effects.

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Keywords: MDR reverter; rat aorta ring; rat tail artery smooth muscle; whole-cell patch-clamp; L-type Ca²⁺ channel inhibitor; dihydropyridines; rhodamine 123; mouse T-lymphoma cell

Abbreviations: BHQ, 2,5-di-*t*-butyl-1,4-benzohydroquinone; DP, dihydropyridine; I_{Ca(L)}, L-type Ca²⁺ current; K60, 60 mM K⁺; MDR, multidrug resistance; MRPs, multidrug resistance-related proteins; P-gp, P-glycoprotein 170; PSS, physiological salt solution; R123, rhodamine 123; TEA, tetraethylammonium; V_h, holding potential; VOCCs, voltage-operated Ca²⁺ channels

Introduction

A large number of drugs are planar lipophilic agents and thus theoretical substrates for the ATP-binding cassette of membrane transporters, including P-glycoprotein 170 (P-gp) and various multidrug resistance (MDR)-related proteins (MRPs), acting as energy-dependent extrusion pumps. The overexpression of these transporters

confers a MDR phenotype on cells in various diseases, including many forms of cancer (Gottesman *et al.*, 2002), rheumatoid arthritis (Llorente *et al.*, 2000), inflammatory bowel disease (Farrell *et al.*, 2000) and epilepsy (Siddiqui *et al.*, 2003).

Development of MDR is one of the main reasons of failure in malignant tumor chemotherapy, as tumor cells, by increasing drug efflux, acquire cross-resistance to many structurally and functionally unrelated anticancer agents,

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which therefore never achieve effective intracellular concentrations (Krishna & Mayer, 2000).

To restore the inherent potency of antitumor agents, several compounds have been studied as inhibitors of P-gp-mediated transport, and then proposed as reverts of cellular resistance (Ford, 1996). So far, MDR reverts belong to various chemical or pharmacological classes including Ca^{2+} channel blockers, calmodulin inhibitors, coronary vasodilators, indole alkaloids, quinolines, hormones, cyclosporines, surfactants and antibodies (Krishna & Mayer, 2000; Kawase & Motohashi, 2003). Among them, Ca^{2+} channel blockers, such as verapamil and some dihydropyridines related to nifedipine, have been extensively investigated (Tsuruo, 1986; Holtt *et al.*, 1992; Toffoli *et al.*, 1995). They are endowed with inherent cardiovascular activity, however, so that the doses required to overcome MDR were found to be associated with severe and sometimes life-threatening cardiovascular toxicity such as atria-ventricular block and hypotension, thus rendering their clinical application impossible. Thus, it is desirable to have compounds that can be used at much lower doses than those shown to be cardiotoxic, or are devoid of cardiovascular effects at the doses effective for reverting MDR. As a consequence, in the last few years, much attention has been focused on the development of safer MDR inhibitors characterized by proper potency, selectivity and pharmacokinetics.

Recently, novel 3,5-diacetyl- (DP1–DP5) and 3,5-dibenzoyl-1,4-dihydropyridine (DP6–DP11) derivatives (Table 1) were synthesized and screened for their ability to reverse MDR in *in vitro* assay systems (Shah *et al.*, 2000; Kawase *et al.*, 2002). Some of them were proved to reduce the activity of P-gp, and therefore they were envisaged as new candidates for MDR reversion in cancer treatment. However, as dihydropyridine derivatives, these compounds are expected to be active also on the cardiovascular system.

The aim of this study, therefore, was to investigate the effects of these dihydropyridines on vascular function *in vitro*,

by comparing their mechanical and electrophysiological actions, as well as to quantify their MDR reversal properties.

Methods

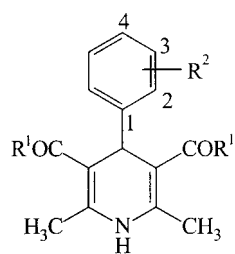
Aorta rings preparation

Aorta rings deprived of endothelium were prepared from male Wistar rats (350–450 g) anesthetized with a mixture of Ketavet® (Gellini, Italy) and Xilor® (BIO 98 srl, Italy), decapitated, and bled. The thoracic aorta was immediately removed, cleaned of adhering fat and connective tissue, and cut into 2.5-mm wide rings with a razor blade slicing device. Each arterial ring was mounted over two rigid parallel stainless steel tubes, one fixed in place and the other attached to an isometric transducer (Basile, Varese, Italy) connected to a kymograph-like pen recorder (Basile, Varese, Italy), or to a flat bed recorder (Linseis, Bolzano, Italy). The preparation was immersed in a water-jacketed organ bath (37°C) containing 5 ml of a modified Krebs–Henseleit physiological salt solution (PSS; see below).

Equilibration period

Rings were allowed to equilibrate for 1 h at a resting tension of 1 g. During this equilibration period, PSS was changed every 15 min and passive tension was re-adjusted to 1 g. Following this equilibration period, rings were stimulated with 60 mM K^+ (K60) PSS, until a sustained response was obtained (~15 min) in order to test their contractile capacity. At the end of a 30-min washout period, the absence of functional endothelium was assessed by the lack of response to 10 μM acetylcholine in rings precontracted with 0.3 μM phenylephrine. Under these conditions, maximal plateau values for active tension of 671.4 ± 14.6 mg ($n = 102$) were obtained. Rings were then washed and equilibrated for another 45-min

Table 1 Structures of the dihydropyridine derivatives tested in the present study



Compounds

Compounds		R^1	R^2
DP1	3,5-Diacetyl-4-(2-nitrophenyl)-1,4-dihydro-2,6-dimethylpyridine	CH_3	2- NO_2
DP2	3,5-Diacetyl-4-(3-phenoxyphenyl)-1,4-dihydro-2,6-dimethylpyridine	CH_3	3-PhO
DP3	3,5-Diacetyl-4-(2-chlorophenyl)-1,4-dihydro-2,6-dimethylpyridine	CH_3	2-Cl
DP4	3,5-Diacetyl-4-(3-chlorophenyl)-1,4-dihydro-2,6-dimethylpyridine	CH_3	3-Cl
DP5	3,5-Diacetyl-4-(4-chlorophenyl)-1,4-dihydro-2,6-dimethylpyridine	CH_3	4-Cl
DP6	3,5-Dibenzoyl-4-(2-nitrophenyl)-1,4-dihydro-2,6-dimethylpyridine	Ph	2- NO_2
DP7	3,5-Dibenzoyl-4-(3-phenoxyphenyl)-1,4-dihydro-2,6-dimethylpyridine	Ph	3-PhO
DP8	3,5-Dibenzoyl-4-(2-chlorophenyl)-1,4-dihydro-2,6-dimethylpyridine	Ph	2-Cl
DP9	3,5-Dibenzoyl-4-(3-chlorophenyl)-1,4-dihydro-2,6-dimethylpyridine	Ph	3-Cl
DP10	3,5-Dibenzoyl-4-(4-chlorophenyl)-1,4-dihydro-2,6-dimethylpyridine	Ph	4-Cl
DP11	3,5-Dibenzoyl-4-phenyl-1,4-dihydro-2,6-dimethylpyridine	Ph	H
Nifedipine	3,5-Dicarbomethoxy-4-(2-nitrophenyl)-1,4-dihydro-2,6-dimethylpyridine	OCH_3	2- NO_2

period before performing the various experimental tests (see below). Control preparations were treated with the vehicle only.

Effects of dihydropyridine (DP) derivatives and nifedipine on rat aorta rings precontracted with K60

The relaxation produced by DP derivatives and nifedipine was assessed in rings precontracted with K60. Cumulative concentration–response curves for each drug were made. Each concentration was left in contact with the ring for long enough to allow full development of the effect. Relaxation was then evaluated as a percentage of the initial tension (taken as 100%).

Assessment of K^+ channel-opening activity

The antispasmodic activity of DP7 was evaluated under different conditions of high K^+ -evoked depolarization (K30 or K60) and compared to the activity of cromakalim (Gurney, 1994). Preparations were stimulated with consecutive additions of K30 or K60, intervalled by washout with PSS, until a stable response was obtained. Following a 45-min washout period, the ring was preincubated with the test drug for 10 min and then stimulated in the presence of the drug, with K30 or K60, respectively. This response was evaluated as a percentage of the response to 0.3 μ M phenylephrine.

Assessment of endoplasmic reticulum Ca^{2+} -mobilizing activity

The effect of DP7 on Ca^{2+} mobilization from the endoplasmic reticulum was evaluated in aortic rings stimulated with phenylephrine, and compared to the activity of 2,5-di-*t*-butyl-1,4-benzohydroquinone (BHQ) (Fusi *et al.*, 1998).

Preparations were incubated with 1 μ M nifedipine. After 15 min, PSS was drained and quickly replaced with K60 containing 1 μ M nifedipine. Under these conditions, K60 failed to contract the rings, demonstrating that extracellular Ca^{2+} influx via L-type Ca^{2+} channels had been blocked. After 5 min, the substances being studied or vehicle were added, for a 15-min incubation period. The rings were then contracted with 1 μ M phenylephrine and the tension expressed as a percentage of the response to 0.3 μ M phenylephrine in PSS. The response to phenylephrine in the presence of nifedipine was taken to represent the contribution of intracellular Ca^{2+} stores to the contractile response. In some experiments, 1 mM Ni^{2+} was added together with nifedipine in order to block Ni^{2+} -sensitive Ca^{2+} channels (see Fusi *et al.*, 1998).

Cell isolation procedure

Smooth muscle cells, freshly isolated from the tail main artery of male rats (350–450 g) by means of collagenase treatment in the presence of bovine serum albumin and trypsin inhibitor, as previously described (Fusi *et al.*, 2001), exhibited an ellipsoid form (10–15 μ m in width, 35–55 μ m in length). The cells were continuously superfused with external solution (see below for composition) using a peristaltic pump (LKB 2132), at a flow rate of 500 μ l min⁻¹. Electrophysiological responses were tested at room temperature (22–24°C) only in those cells that were

phase dense. Cell membrane capacitance averaged out at 48.55 ± 1.52 pF ($n = 50$).

Whole-cell patch-clamp recording

Conventional whole-cell patch-clamp method (Hamill *et al.*, 1981) was employed to voltage-clamp the smooth muscle cells. Recording electrodes were pulled from borosilicate glass capillaries (WPI, Berlin, Germany) and fire-polished to give a pipette resistance of 2–5 M Ω when filled with internal solution (see below). A low-noise, high-performance Axopatch 200B (Axon Instruments, U.S.A.) patch-clamp amplifier, driven by an IBM computer in conjunction with an A/D, D/A board (DigiData 1200 A/B series interface, Axon Instruments, U.S.A.), was used to generate and apply voltage pulses to the clamped cells and record the corresponding membrane currents. Current signals, after compensation for whole-cell capacitance, series resistance and liquid junction potential, were low-pass filtered at 1 kHz and digitized at 3 kHz prior to being stored on the computer hard disk. Long-lasting nifedipine-blockable inward currents passing through L-type Ca^{2+} channels [$I_{Ca(L)}$], in 5 mM Ca^{2+} -containing external solution (see below), were elicited with 250-ms depolarizing voltage-clamp pulses to test potentials of 10 mV from a holding potential (V_h) of –50 mV. Data were collected once the current amplitude had been stabilized (usually 8–10 min after the whole-cell configuration had been obtained). $I_{Ca(L)}$ did not run down during the following 30–40 min under these conditions.

K^+ currents were blocked with 30 mM tetraethylammonium (TEA) in the external solution and Cs^+ in the internal solution.

Current values were corrected for leakage using 1 μ M nifedipine, which was assumed to completely block $I_{Ca(L)}$.

Rhodamine 123 (R123) uptake assay

The L5178 Y mouse T-lymphoma parent cell line was transfected with a recombinant MDR1/A retroviral vector (pHa MDR1/A), as previously described by Pastan *et al.* (1988). MDR1-expressing cell lines were selected by culturing the infected cells with 60 ng ml⁻¹ colchicine to maintain the expression of the MDR phenotype (Weaver *et al.*, 1993). The L5178 MDR cell line and the L5178 Y parent cell line were grown in McCoy's 5A medium supplemented with 10% heat-inactivated horse serum, 2 mM L-glutamine and 20 μ g ml⁻¹ penicillin, 20 μ g ml⁻¹ streptomycin and 8300 U l⁻¹ nystatin.

Cells (2×10^6 ml⁻¹) were resuspended in serum-free McCoy's 5A medium, and 0.5 ml aliquot of the cell suspension was distributed into each Eppendorf centrifuge tube. Compounds to be tested were added at different concentrations and samples were incubated for 10 min at room temperature. Then, R123 indicator was added to the samples at a final concentration of 5.2 μ M and cells were incubated for 20 min at 37°C, washed twice and resuspended in 0.5 ml phosphate-buffered saline. The uptake of R123 by cells was quantified as fluorescence by flow cytometry, using a FACScan instrument (Beckton Dickinson, Oxford, U.K.) at 488 nm and 300 mW. In order to measure P-gp inhibition in cells treated with DP derivatives, the fluorescence intensity of treated cells/fluorescence intensity of untreated cells ratio was transformed into its reciprocal and multiplied by 100. The percent residual of

fluorescence intensity, thus obtained, was subtracted from 100, to give the percent P-gp inhibition value.

Solutions and chemicals

PSS contained (in mM) 118 NaCl, 4.75 KCl, 2.5 CaCl₂, 1.19 MgCl₂, 1.19 KH₂PO₄, 25 NaHCO₃ and 11.5 glucose, bubbled with a 95% O₂–5% CO₂ gas mixture to give a pH of 7.4. PSS containing KCl at a concentration of more than 4.75 mM was prepared by replacing NaCl with equimolar KCl.

Ca²⁺-free external solution contained (in mM): 110 NaCl; 5.6 KCl; 10 HEPES; 20 taurine; 20 glucose; 1.2 MgCl₂ and 5 Na-pyruvate (pH 7.4).

Internal solution (pCa 8.4) consisted of (in mM): 100 CsCl, 10 HEPES, 11 EGTA, 2 MgCl₂, 1 CaCl₂, 5 Na-pyruvate, 5 succinic acid, 5 oxalacetic acid, 3 Na₂ATP and 5 phosphocreatine; pH was adjusted to 7.4 with CsOH.

The osmolarity of external solution was adjusted to 335 mosm, and that of the internal solution to 310 mosm (Stansfeld & Mathie, 1993), by means of an osmometer (Osmostat OM 6020, Menarini Diagnostics, Italy).

The chemicals used were: collagenase (type XI), TEA, bovine serum albumin, trypsin inhibitor, verapamil, nifedipine, BHQ, cromakalim, colchicine, horse serum, L-glutamine, penicillin, streptomycin, nystatine, McCoy's 5A medium, phosphate-buffered saline and R123 (Sigma Chimica, Italy).

DP1–DP5 and DP6–DP11 were synthesized by Shah *et al.* (2000) and Kawase *et al.* (2002), respectively.

Nifedipine dissolved directly in ethanol, and cromakalim, BHQ and DP derivatives, dissolved in DMSO, were diluted in PSS, external solution or McCoy's 5A medium, prior to use. The resulting concentrations of DMSO and ethanol (below 1 or 0.55% for aorta rings, 0.3 or 0.1% for tail myocytes and 0.5 or 1% for L5178 MDR cell line, respectively) did not affect the responses (data not shown).

Statistical analysis

Acquisition and analysis of data were accomplished using pClamp 8.2.0.232 software (Axon Instruments, U.S.A.) and GraphPad Prism version 3.03 (GraphPad Software, U.S.A.). Data are reported as mean \pm s.e.m.; *n* represents the number of aorta rings or tail myocytes, isolated from at least three animals.

Statistical analyses and significance, as measured by ANOVA, followed by Dunnett's post test, were obtained using GraphPad InStat version 3.05 (GraphPad Software, U.S.A.). In all comparisons, *P* < 0.05 was considered significant.

Results

Effects of DP derivatives and nifedipine on contraction promoted by extracellular Ca²⁺ influx at K60

In this set of experiments, muscle tension developed at K60 was taken as an index of Ca²⁺ influx from the extracellular space promoted by membrane depolarization. The effect of DP derivatives is summarized in Figure 1b. DP4 was the most potent myorelaxing agent (Figure 1a,b), although characterized by an IC₅₀ two orders of magnitude higher than that of nifedipine (Table 2). DP6, DP8, DP1 and DP2 exhibited IC₅₀ values one order of magnitude higher than that of DP4,

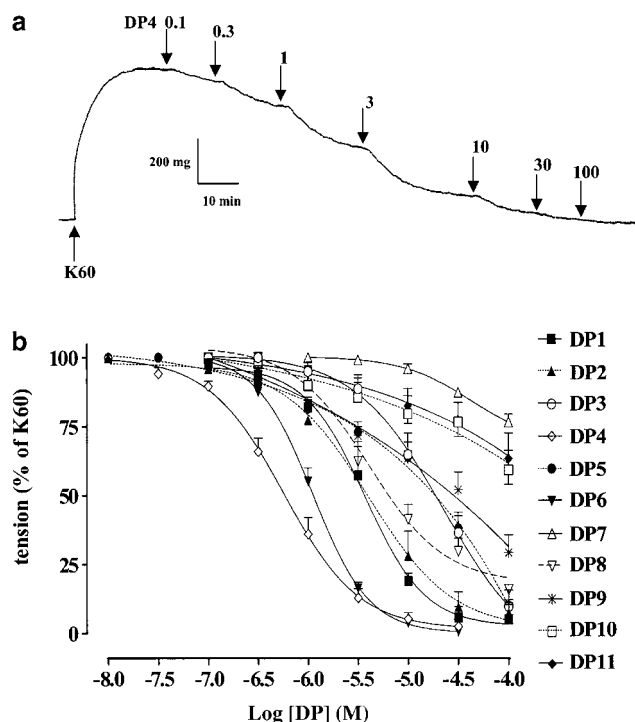


Figure 1 Effect of DP derivatives on Ca²⁺-induced contraction in rat aorta rings depolarized with K60. (a) Trace (representative of at least four experiments) of relaxation developed in response to cumulative concentrations (μ M) of DP4 added on the plateau of K60-elicited contraction. (b) Concentration–response curves for DP derivatives. Data points are mean \pm s.e.m. (*n* = 3–6).

Table 2 Inhibition of Ca²⁺-induced contractions by dihydropyridine derivatives in rat aorta rings depolarised with K60 (figures are means \pm s.e.m. (*n* = 3–6))

Compounds	IC ₅₀ (M)
DP1	$3.69 \pm 0.03 \times 10^{-6}$
DP2	$3.97 \pm 0.04 \times 10^{-6}$
DP3	$2.23 \pm 0.03 \times 10^{-5}$
DP4	$5.65 \pm 0.02 \times 10^{-7}$
DP5	$1.39 \pm 0.02 \times 10^{-5}$
DP6	$1.11 \pm 0.01 \times 10^{-6}$
DP7 ^a	$> 3 \times 10^{-5}$
DP8	$4.04 \pm 0.08 \times 10^{-6}$
DP9 ^a	$> 3 \times 10^{-5}$
DP10 ^a	$> 3 \times 10^{-5}$
DP11 ^a	$> 3 \times 10^{-5}$
Nifedipine	$1.72 \pm 0.03 \times 10^{-9}$

^aFigures reported refer to the highest concentration tested, at which the effect observed did not exceed 50%.

although DP8 at the maximum concentration tested caused a spasmodic relaxation that did not exceed 70.4%. DP5 and DP3 IC₅₀ values were about two orders of magnitude higher than that of DP4. The compounds endowed with the lowest spasmodic activity (DP9, DP10, DP11 and DP7 up to 30 μ M) relaxed K60-induced contraction by less than 50%.

Effects of DP derivatives and nifedipine on I_{Ca(L)}

All the novel dihydropyridine derivatives, but DP7, inhibited I_{Ca(L)} in a concentration-dependent manner (Figure 2b). As shown in Table 3, DP6 was the most potent I_{Ca(L)} inhibitor

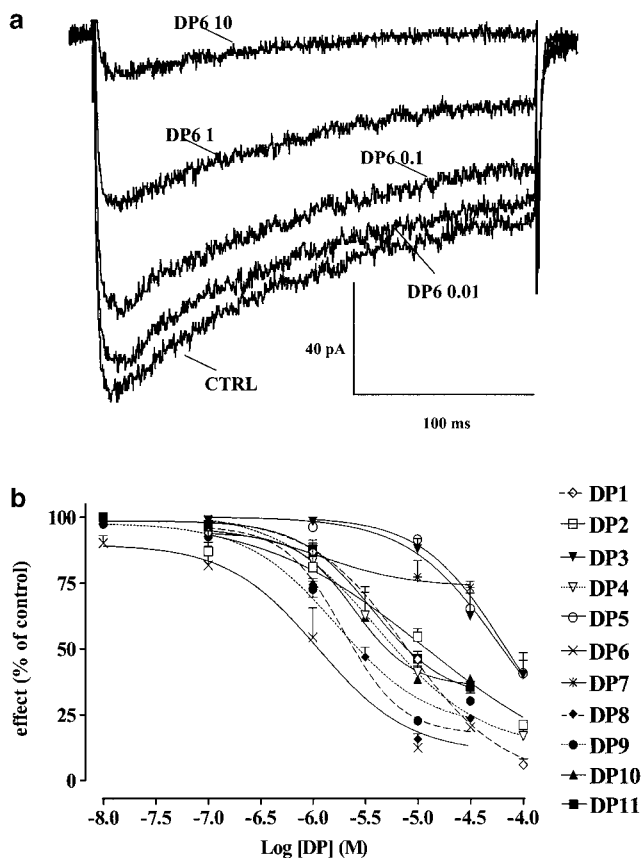


Figure 2 Effect of DP derivatives on $I_{Ca(L)}$ recorded in rat tail myocytes. (a) Original recordings of conventional whole-cell $I_{Ca(L)}$ elicited with 250-ms clamp pulses to 10 mV from a V_h of -50 mV, measured in the absence (CTRL) or presence of DP6 (0.01–10 μ M). (b) Concentration–response curves for DP derivatives. Data points are mean \pm s.e.m. ($n = 3$ –6).

Table 3 Inhibition of $I_{Ca(L)}$ by dihydropyridine derivatives in rat tail artery myocytes (figures are means \pm s.e.m. ($n = 3$ –6))

Compounds	IC_{50} (M)
DP1	$7.93 \pm 0.08 \times 10^{-6}$
DP2	$1.21 \pm 0.03 \times 10^{-5}$
DP3	$6.35 \pm 0.05 \times 10^{-5}$
DP4	$4.34 \pm 0.02 \times 10^{-6}$
DP5	$6.90 \pm 0.07 \times 10^{-5}$
DP6	$1.12 \pm 0.04 \times 10^{-6}$
DP7 ^a	$> 3 \times 10^{-5}$
DP8	$1.95 \pm 0.06 \times 10^{-6}$
DP9	$1.71 \pm 0.08 \times 10^{-6}$
DP10	$2.48 \pm 0.05 \times 10^{-6}$
DP11	$3.71 \pm 0.03 \times 10^{-6}$
Nifedipine	$4.48 \pm 0.07 \times 10^{-9}$

^aFigure reported refers to the highest concentration tested, at which the effect observed did not exceed 20%.

(Figure 2a), although its IC_{50} value was three orders of magnitude higher than that of nifedipine.

DP1, DP4, DP8, DP9, DP10 and DP11 also exhibited remarkable Ca^{2+} antagonist properties and showed IC_{50} values ranging between 1.95 and 7.93×10^{-6} , while DP2, DP3 and DP5 inhibited $I_{Ca(L)}$ with IC_{50} values one order of magnitude higher than that of DP6.

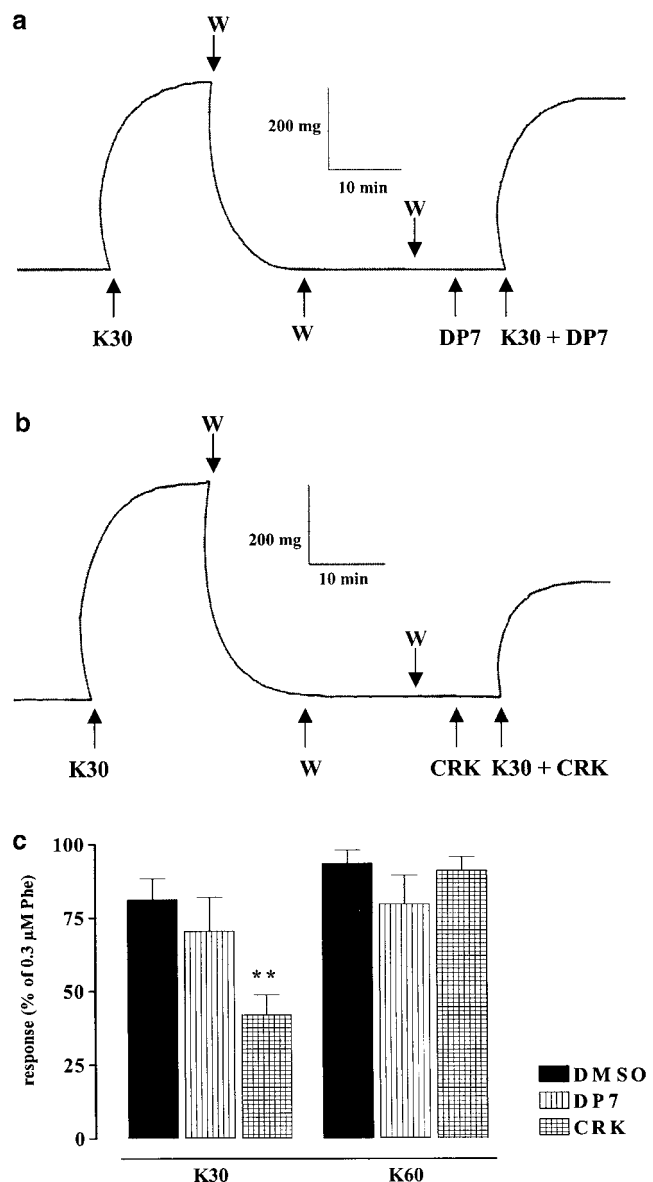


Figure 3 Effect of DP7 and cromakalim (CRK) on Ca^{2+} -induced contractions in rat aorta rings depolarized with either K30 or K60. Rings were bathed with K30 for 15 min and then washed in PSS, until a stable response was obtained. DP7 (30 μ M) (a) or CRK (500 nM) (b) were added 5 min before K30. Traces are representative of at least four experiments. (c) Rings were stimulated with either K30 or K60 in the presence of DMSO (vehicle), 30 μ M DP7 or 500 nM CRK. Columns represent mean \pm s.e.m. ($n = 4$ –8). ** $P < 0.01$ (Dunnett's post-test).

Effects of DP7 and cromakalim on contractions promoted by extracellular Ca^{2+} influx at either K30 or K60

K^+ channel-opening activity was assessed by stimulating aorta rings with K30 (Figure 3a, b) or K60 in the presence of the tested compounds. K30 and K60 evoked contractile responses averaging $81.1 \pm 7.2\%$ ($n = 6$) and $93.5 \pm 4.5\%$ ($n = 8$), respectively, of the maximum attainable contraction (Figure 3c).

In rings preincubated with 30 μ M DP7, both K30- and K60-induced contractions were not significantly different from control. In contrast, the relaxant effect of cromakalim, a K^+ channel opener, could be observed at K30 but not at K60.

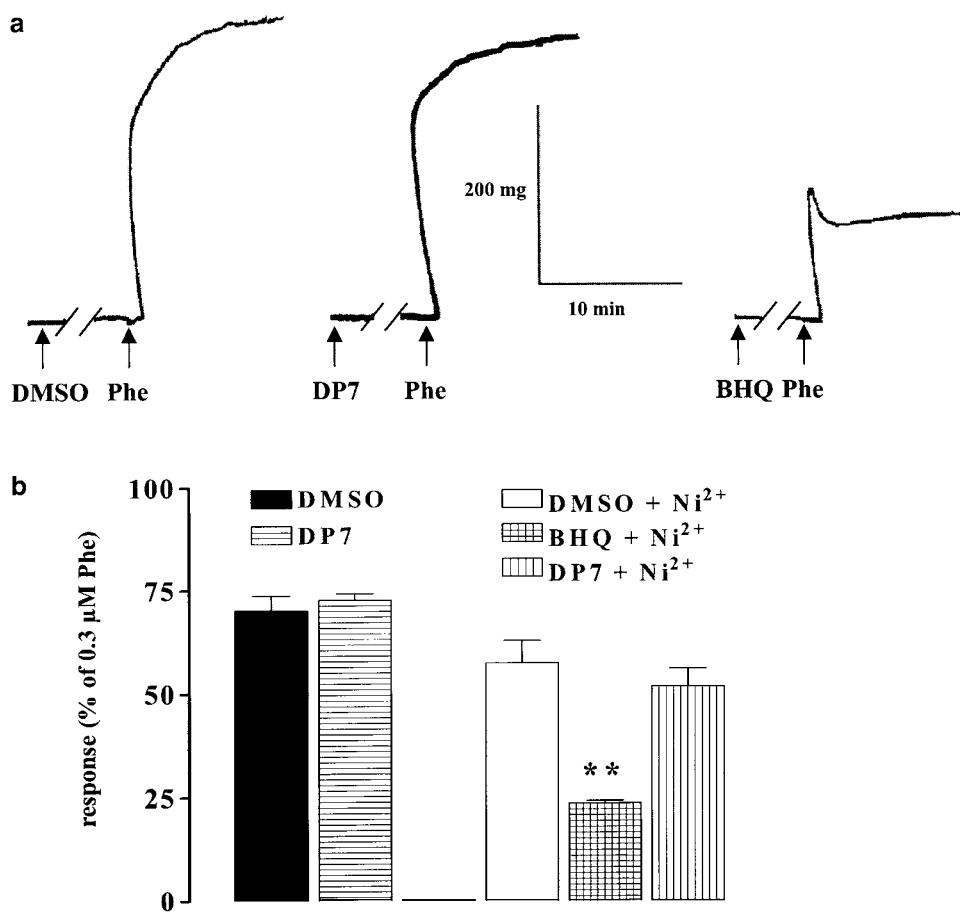


Figure 4 Effect of DP7 and BHQ on Ca^{2+} mobilization from intracellular stores. All traces show $1 \mu\text{M}$ phenylephrine (Phe)-induced contraction of rat aorta rings incubated in K60 containing $1 \mu\text{M}$ nifedipine and 1 mM Ni^{2+} . This response was assumed to represent the contribution of intracellular Ca^{2+} sources to the contractile response. (a) Phe-induced response after 10 min incubation with DMSO (vehicle), with $30 \mu\text{M}$ DP7, or with $10 \mu\text{M}$ BHQ. Traces are representative of at least four experiments. (b) Effect of DP7 and BHQ on Phe-induced contraction in K60 containing nifedipine or nifedipine plus Ni^{2+} . Columns represent mean \pm s.e.m. ($n = 4$). $**P < 0.01$ (Dunnett's post-test).

Effects of DP7 and BHQ on contraction promoted by Ca^{2+} mobilization from the endoplasmic reticulum

The amount of Ca^{2+} stored intracellularly was estimated from the amplitude of the phenylephrine-induced contraction in the presence of K60 and nifedipine or nifedipine plus Ni^{2+} . Figure 4a shows the typical traces of $1 \mu\text{M}$ phenylephrine-induced contraction in rings depolarized with K60 in the presence of nifedipine plus Ni^{2+} with or without $30 \mu\text{M}$ DP7 or $10 \mu\text{M}$ BHQ. In the presence of nifedipine or nifedipine plus Ni^{2+} , phenylephrine evoked tonic responses (70.1 ± 3.7 and $57.5 \pm 5.4\%$, respectively, of the maximum attainable contractions, $n = 4$), which were not affected by the presence of DP7 (Figure 4b). On the contrary, when tissues were preincubated with BHQ, a quite selective endoplasmic reticulum Ca^{2+} -ATPase inhibitor (Moore *et al.*, 1987), the response to phenylephrine was significantly reduced and characterized by an initially phasic component, followed by a tonic contraction.

Effects of DP derivatives and nifedipine on P-gp-mediated R123 efflux

All the novel dihydropyridine derivatives, but DP1, DP2 and DP4, inhibited P-gp-mediated R123 efflux in a concentration-

dependent manner (Table 4). DP7 was the most potent P-gp inhibitor with an IC_{50} value two orders of magnitude lower than that of nifedipine. DP9 and DP10 showed IC_{50} values comparable to that of DP7. DP6, DP8 and DP11 also exhibited a remarkable MDR-reversing activity, and showed IC_{50} values ranging between 2.74 and 7.35×10^{-6} , while DP3 and DP5 were as active as nifedipine.

DP1, DP2 and DP4, at the maximum concentration tested ($30 \mu\text{M}$ for DP1 and DP2, and $165 \mu\text{M}$ for DP4), inhibited P-gp activity by only 30%.

Discussion

Endeavouring to find MDR reverters is a crucial task for anticancer chemotherapeutic intervention. In the past few years, extensive studies have been performed with the aim of developing effective chemosensitizers to overcome MDR of human cancer cells. Potent P-gp inhibitors have been tested in clinical trials so far, including Ca^{2+} channel blockers such as verapamil and dihydropyridines (Bates *et al.*, 1994; Bellamy, 1996). However, clinical application of these agents has not been extensively pursued to date, owing to their unwanted cardiovascular side effects. This has promoted the search for

Table 4 Inhibition of Pgp-mediated R123 efflux by dihydropyridine derivatives in L5178 MDR cell line (figures are means \pm s.e.m. from triplicate experiments)

Compounds	IC ₅₀ (M)
DP1 ^a	$> 3 \times 10^{-5}$
DP2 ^a	$> 3 \times 10^{-5}$
DP3	$4.27 \pm 0.19 \times 10^{-5}$
DP4 ^a	$> 16.5 \times 10^{-5}$
DP5	$2.15 \pm 0.06 \times 10^{-5}$
DP6	$2.74 \pm 0.07 \times 10^{-6}$
DP7	$3.02 \pm 0.07 \times 10^{-7}$
DP8	$5.92 \pm 0.11 \times 10^{-6}$
DP9	$6.90 \pm 0.05 \times 10^{-7}$
DP10	$7.07 \pm 0.19 \times 10^{-7}$
DP11	$7.35 \pm 0.10 \times 10^{-6}$
Nifedipine	$2.01 \pm 0.04 \times 10^{-5}$

^aFigures reported refer to the highest concentration tested, at which the effect observed did not exceed 30%.

congeners of this first-generation-MDR reverters with reduced cardiovascular toxicity.

A series of 3,5-diacetyl- (DP1–DP5) and 3,5-dibenzoyl-1,4-dihydropyridine (DP6–DP11) derivatives (Table 1), recently synthesized and preliminarily assessed for their ability to reverse MDR in *in vitro* assay systems, have revealed a noteworthy P-gp inhibitory activity in L5178 Y mouse T-lymphoma cells transfected with MDR1 gene (Shah *et al.*, 2000; Kawase *et al.*, 2002). In order to provide further information on these new candidates for MDR cancer cell treatment, their effects on vascular functions *in vitro* have been investigated and compared to their MDR-reversing potency. All compounds tested, but DP9, DP10, DP11 and DP7, antagonized K⁺-induced contraction in a concentration-dependent manner similar to the well-known Ca²⁺-channel-blocking drug nifedipine. K⁺-induced contraction is the result of an increased Ca²⁺ influx through voltage-operated Ca²⁺ channels (VOCCs). Therefore, its inhibition by the DP derivatives might be interpreted as a consequence of the blockade of these channels. The electrophysiological data presented here, in fact, have directly confirmed this hypothesis, since all dihydropyridines tested, but DP7, inhibited, although to a different extent, *I*_{Ca(L)} recorded in rat tail myocytes.

Comparison of the Ca²⁺ antagonist activity between DP compounds and nifedipine has shown that the latter was about 1000-fold more potent in both model systems. This could be due to the replacement of the 3,5-dicarbomethoxy groups of nifedipine by either 3,5-diacetyl (DP1–DP5) or 3,5-dibenzoyl (DP6–DP11) groups. This is in agreement with what previously reported by Loev *et al.* (1974) and Janis & Triggle (1983), who showed that replacement of 3,5-dicarbomethoxy groups by other electron-withdrawing substituents markedly reduces the Ca²⁺-blocking activity of the resulting compounds.

DP1, DP3, DP5, DP6 and DP8 showed an approximately equivalent activity in both model systems. In contrast, DP9, DP10 and DP11 were more effective in inhibiting *I*_{Ca(L)} in single smooth muscle cell than in relaxing aorta rings, suggesting that a different diffusion of the compounds across intact tissues and isolated myocytes might account for this difference. However, this feature alone cannot account for their lower effectiveness towards K⁺-induced contraction,

since DP9, DP10 and DP11 are equally lipophilic to DP6 and DP8 (Shah *et al.*, 2000; Kawase *et al.*, 2002). It is likely that the position of the substituent in the 4-phenyl ring may be critical: para (DP10) and meta (DP9 and DP11) substitution invariably decreased the spasmolytic activity, whereas ortho (DP6, DP8) substitution increased it. Moreover, slight differences in L-type Ca²⁺ channels expressed in the two tissues examined might explain the incomplete matching of data from the two experimental settings. Finally, DP2 and DP4 caused a much higher spasmolysis than what could be expected from their Ca²⁺ channel-blocking activity, thus advocating their interaction with targets, beyond VOCCs, crucial for the maintenance of vascular smooth muscle tone.

Noteworthy, among the seven derivatives (DP1, DP2, DP3, DP4, DP5, DP6 and DP8) exhibiting both spasmolytic and Ca²⁺ channel-blocking activity, a linear and significant correlation was found ($r^2 = 0.65$, $P < 0.05$), thus indicating that homologies between the two vascular preparations employed in this study are more relevant than differences.

Safak *et al.* (1990) have already tested the Ca²⁺ antagonist activity of DP1 and DP3 in K⁺-depolarized guinea-pig *Taenia coli* strips. In this preparation, DP1 and DP3 showed IC₅₀ values one order of magnitude lower than those reported here. The higher concentration of extracellular Ca²⁺ used in the present work (2.5 mM *versus* 1 mM in *Taenia coli* strips) might account for this difference, since the extent of inhibition of muscle contraction by Ca²⁺ channel-blocking agents is known to be inversely correlated to the extracellular Ca²⁺ concentration (see Karaki, 1987).

Since DP7 was almost ineffective as a vascular Ca²⁺ channel blocker, its effects on other functions that regulate vascular smooth muscle tone were assessed.

The antispasmodic activity of K⁺ channel openers, such as cromakalim (Norman *et al.*, 1994), is reduced when extracellular K⁺ concentration is raised; invariably, when the K⁺ gradient across the membrane drops, the effect of K⁺ channel openers disappears (Gurney, 1994; this paper). In contrast to what observed with cromakalim, however, DP7 did not show any antispasmodic activity even when the external K⁺ concentration was kept at a value (30 mM) which allows K⁺ channel activation to be displayed, nor did it affect muscle contraction caused by intracellular stored Ca²⁺ mobilization.

In conclusion, DP7, at a concentration two orders of magnitude higher than IC₅₀ as a P-gp inhibitor, was neither an antispasmodic nor a spasmolytic agent and inhibited *I*_{Ca(L)} by a mere 20%, thus outlining the clear divergence of its P-gp inhibition and the vascular activity. The absence of vascular activity *per se*, however, is not sufficient to guarantee for DP7 safety, and further experiments on other physiological systems are needed before it can be addressed to the clinical investigation. Nevertheless, since reversion of MDR is a field of research of overwhelming importance, DP7 may represent a lead compound for the design of novel, safe and potent MDR chemosensitizers needed for the chemotherapy of cancer and other diseases.

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